

## Effect of cement dust deposition on physiological behaviors of some halophytes in the salt marshes of Red Sea

Amal M. Abdel- Rahman<sup>1,2</sup> and Mohamed M. Ibrahim<sup>1,3</sup>

1-Alexandria University, Faculty of Science- Botany and Microbiology department  
P.O. Box. 21511; Alexandria. EGYPT

2-Umm Al Qura University, Faculty of Education for Girls, Makkah, K.S.A.

3-King Saud University, Teachers College; Science department; P.O. Box 4341  
Riyadh 11491, K.S.A.

Corresponding author: [m\\_ibramim2004@yahoo.com](mailto:m_ibramim2004@yahoo.com)

### ABSTRACT

Deposition effect of cement dust on pigmentation was studied in *Zygothallum coccineum*, *Salsola tetrandra*, *Cyperus conglomeratus*, *Limonium axillare* and *Suaeda vermiculata*. Measurements were taken for plants in two sites, one close to early constructed cement factory at (Jeddah, Saudi Arabia) about 500 meters and other one far from the factory about 5 kilometers. The rate of mortality of young branches were high in the area subjected to the cement dust in all five species, whereas *Salsola tetrandra* and *Suaeda vermiculata* are the most sensitive among them. Unless *Cyperus conglomeratus*, It is noted that amount of chl a, b and carotenoids in all investigated plants that are far away from cement dust more than that near from the factory. Carbohydrates and protein contents are significantly decreased in non polluted *Salsola tetrandra* and *Suaeda vermiculata*, whereas these parameters increased significantly parallel to the dust accumulation rate on plants near to the factory. In general, pollution by the cement dust has caused adverse effects on the photosynthetic pigments, the pH of the cell sap, metabolism of soluble amino acids and soluble sugars.

**Key words:** Halophytic species – salt marshes- Cement dust- Photosynthetic pigments.

### INTRODUCTION

Physiological disorders such as reduced growth is ultimately due to the cumulative effects of the causal factors on the physiological processes necessary for plant growth and its development (Schutzki and Cregg, 2007). Due to immobility of higher plants, it need a greater protection against several stresses, including low and high temperature, water stress, salinity, metal toxicity, and others. Air pollution has a diverse effect on many metabolic process in plants such as photosynthetic activities, mitochondrial respiration and stomatal clogging of plants (Miller *et al.*, 1973). Over the last three decades, the industrial and human activities in the coastal area of Kingdom of Saudi Arabia have increased dramatically and resulted in the continuous invasion of different types of pollutants including heavy metals (Holick, 1995). Bader *et al.* (2009) studied three coastal areas along the Red Sea of Kingdom of Saudi Arabia and they concluded that Pollution Load Index (PLI) was significantly high in Jeddah and it was considered as the most polluted area, followed by Rabigh while Yanbu was the least contaminated area. Industries are emitting toxic substances which adversely affect man's food supply by polluting nearby growing plants. Ambient level of air pollution has been shown to affect stomatal conductance, photosynthesis and root morphology of young beech (Taylor and Davies, 1990). One

of the most recent studies of these stresses was a dust accumulation, which provokes severe damage in the photosynthetic apparatus (Santosh and Tripathi, 2008). Photosynthesis is known to be one of the most stress-sensitive processes and it can be completely inhibited by stress before other symptoms of the stress are detected (Berry and Bjorkman, 1980). Different cements have different ingredients; many of them contain substances that can be hazardous, like crystalline silica (quartz), lime, gypsum, nickel, cobalt, and chromium compounds (Green, 2004). Wind erosion suspends large quantities of dust in the atmosphere that settle back to the earth's surface and are deposited on plant leaves when wind velocities decrease (Armbrust, 1986). Cement factories are major source of pollutants for the surrounding areas (Stratmann and Van Haut, 1966). Dusted plants with quantities of dust deposition ranging from 1 to 48 g/m<sup>2</sup> per day; dust falling on the soil caused a shift in the pH level to the alkaline side. It was found that dust deposition affect photosynthesis, stomatal functioning and productivity (Santosh and Tripathi, 2008). A scant numbers of literatures showed that there is a relation between cement dust deposition and physiological process in halophytic weed plants. Since the studying area is suffering from heavy cement dust which deposits on the buildings and plants and producing a significant adverse effect. So that, our goal in the present study was to evaluate the relationships between cement dust deposition and various physiological parameters of five halophytic species; *Zygophyllum coccineum*, *Salsola tetandra*, *Cyperus conglomeratus*, *Limonium axillare* and *Suaeda vermiculata*. Based on the results of these physiological parameters, insight into the mechanism responsible for dust tolerance in different plants will be elucidated.

## MATERIAL AND METHODS

The study area was chosen in Rabigh which located along the Red Sea coast 165 Km north Jeddah, (Fig. 1). Vegetation of this area is suffering from dust pollution due to the presence of cement factory. Five dominants halophytic species, *Zygophyllum coccineum*, *Salsola tetandra*, *Cyperus conglomeratus*, *Limonium axillare* and *Suaeda vermiculata* were selected at two different locations down-wind from a cement factory in the studying area. Location (1, 2) were about 450-500 m and 4500-5000 m from the factory respectively. Selected plant species and soil samples were collected randomly from the two locations covering the study area in the season of autumn.

**Soil analysis:** Soil samples were collected from adjacent plants samples at a depth from 0-25 cm. Physical and chemical parameters of such soil samples were analyzed. Soil (pH was determined in the soil paste according to the method described by Kilmer and Alexander (1949) by using pH-meter (4010 data. Hani. Instruments, Padova, Italy). Electrical conductivity (EC) of the saturated soil extracts were determined as described by Kilmer and Alexander (1949) and expressed as dSm<sup>-1</sup>. The anions and cations (Cl, SO<sub>4</sub>, Ca, Mg, Na and K) of the soil extracts were analyzed as described by Richards, 1954; Jacobson and Hill, 1970 and their values expressed as meq L<sup>-1</sup>. The chemical and physical analyses of the dust collected in the vicinity of the Cement Factory of Rabigh were carried out.

Leaves were prepared and analyzed for the determination of photosynthetic pigments according to Metzner *et al.*(1965), carbohydrates in dry leaves were determined according to Naguib (1964) and Murata (1968); soluble and total protein were determined in dry leaves according to Hartree (1972) and some elements (Mg, Fe, Mn, N) were also determined according to Allen (1974). Electrolyte leakage and

pH values were measured by using conductivity meter (4010 data. Hani. Instruments, Padova, Italy) according to method described by Dicagno *et al.*(1999). Leaf disks (1cm) were collected and washed with de-ionized water to remove surface-adhered electrolytes and incubated overnight at 25°C, with gently shaken on a gyratory shaker.

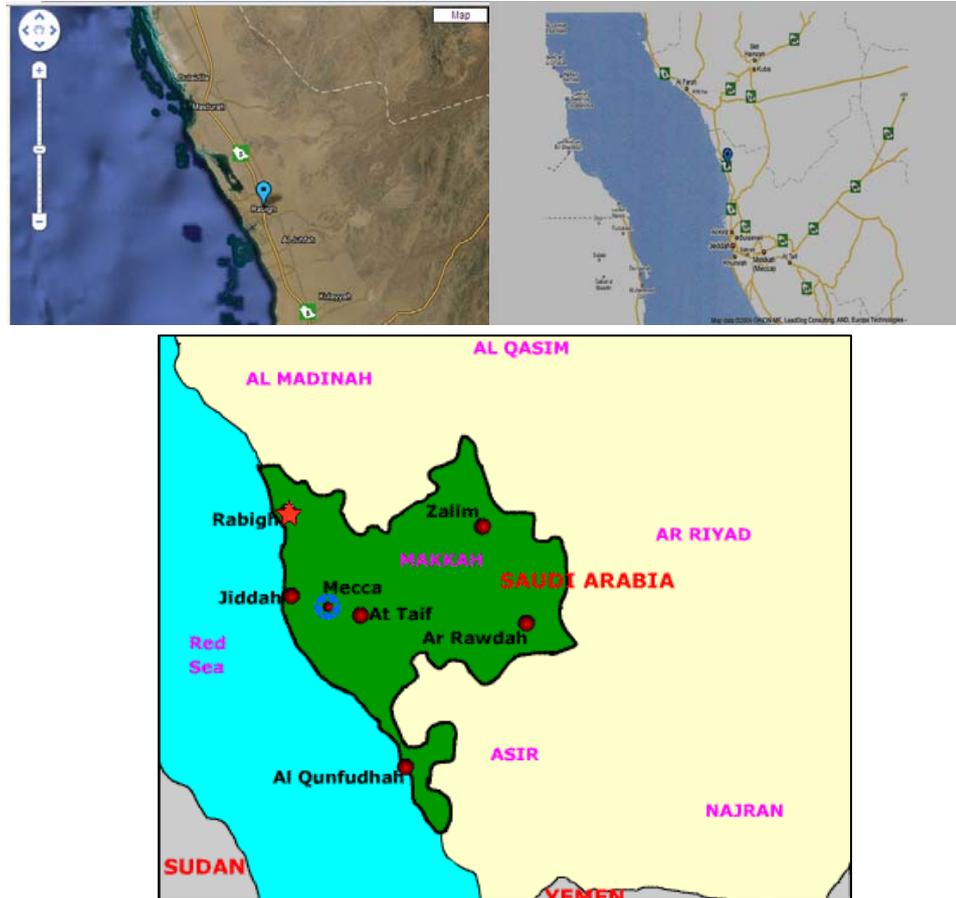


Fig. 1: Map showing the study location

[http://www.mapzones.com/maps/saudi\\_arabia/makkah/rabigh.php](http://www.mapzones.com/maps/saudi_arabia/makkah/rabigh.php)

After measurements, the tubes were then placed in a boiling water for 15 min, and then cooled to room temperature and shaken for 2h .Conductivity was again determined .The electrolyte leakage was calculated as the ratio of conductivity before boiling to that after boiling.

#### Analysis of cement

The determination of a wide range of elements in various samples of cement was performed using atomic absorption according to Thomas (1969).

### RESULTS

The elemental profile, determined in the soil extract of studied plants for the two locations, indicated that the level of calcium, magnesium, sodium, potassium and sulphate were significantly higher in the polluted soil than in the control one (Fig. 2). Analysis of the dust collected from the leaf's surface indicated the presence of some toxic heavy metals such as aluminum silicon and cadmium, in addition to some other elements (Table 1). Dust accumulation altered the chlorophyll and carotenoid contents in all plants in the polluted location (near the factory) compared with plants far from

the factory in control site (). Greater decrease in chlorophyll a and b contents were clearly observed in *Salsola tetandra* and *Limonium axillare* (For chl a about -36% and -45%, respectively). A similar pattern of decrease was occurred in *Zygophyllum coccineum* and *Suaeda vermiculata* but to lower extent (about -15 and -9%, respectively).

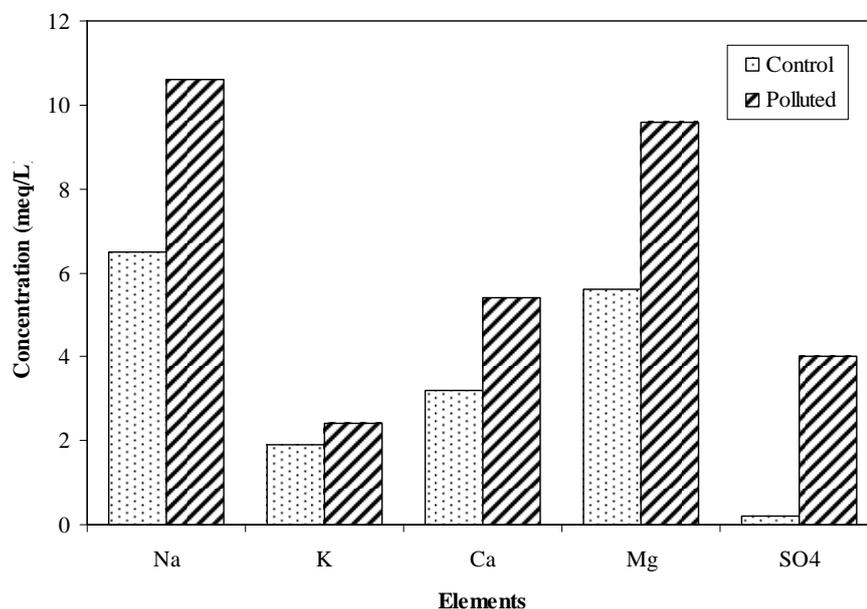


Fig. 2: Concentration of Na, K, Ca, Mg and SO<sub>4</sub> in soil of the studied areas, location1 far from the cement factory (control), location 2 near to the cement factory (polluted). Values significantly different from non polluted in the same plant type according to F-test ( $p < 0.05$ ).

Table 1: Analysis of element in dust collected around the Cement Factory from plant studied and soil components in study area

Component	Atomic Absorption sample (%)	Soil component	%
SiO <sub>2</sub>	22.49	Maximum water holding capacity	65.72%
Al <sub>2</sub> O <sub>3</sub>	4.20	Soil pH	9.53
Fe <sub>2</sub> O <sub>3</sub>	2.58	Soil texture	Clay loam
CaO	65.8	Calcium carbonate	22%
MgO	1.25	Alkaline carbonate	2.45 meq/l
Na <sub>2</sub> O	0.13	Amount of chloride	8.6 meq/l
K <sub>2</sub> O	0.29	Conductivity	506 $\mu$ s/cm

On the other hand, Chlorophyll a, b and chl a/b ratio were significantly increased in *Cyperus conglomeratus* together with increasing dust accumulation in plant leaves near to the factory (Table2 and Fig.3). Chlorophyll a/b ratio for the other four studied plants were decreased in the polluted plant's leaves near the factory and compared with plants far from the dust. Also, the change in the carotenoids content showed the same pattern. There was a significant reduction in carotenoids content particularly for *Suaeda vermiculata* *Salsola tetrandra*, and *Limonium axillare* to less than 10, 33, and 46 %, respectively (Fig.3). For the leaves of *Zygophyllum coccineum* and *Cyperus conglomeratus* a substantial increase in the carotenoids content in response to dust accumulation near the factory occurred. Thus 42 % and 23% of the increase in the content compared to the control plants, respectively. The chlorophyll/carotenoid ratio decreased in *Zygophyllum coccineum* and *Salsola*

*tetandra* plants near to the factory indicating the increase in carotenoid content with respect to non polluted plants (Table2).

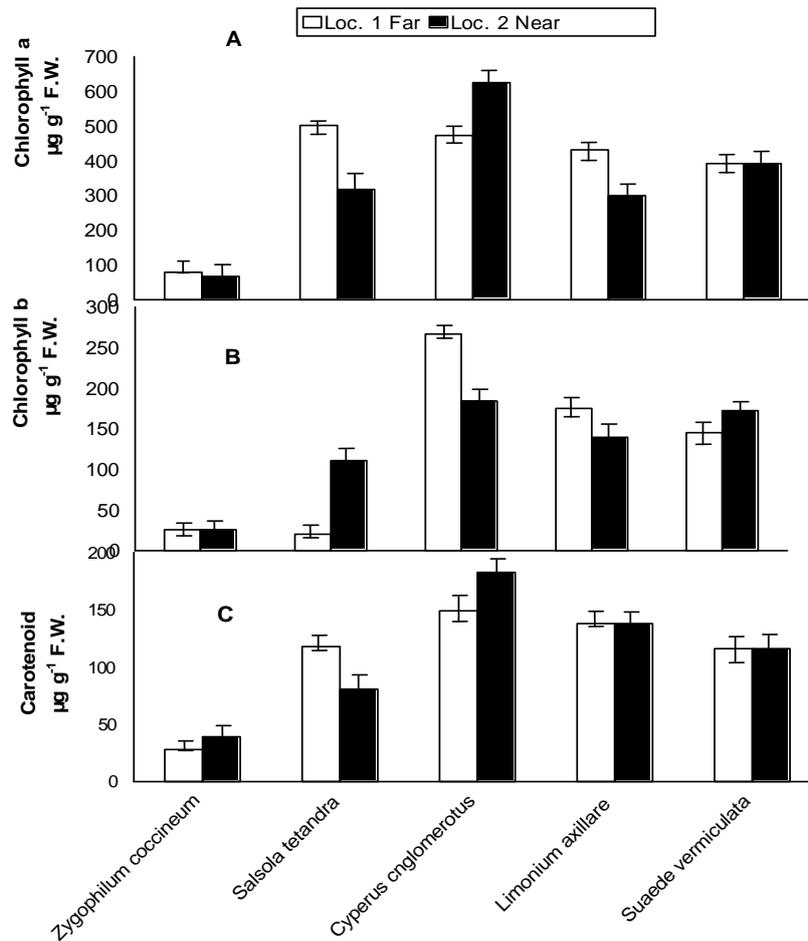


Fig. 3: Chlorophyll a (A), chlorophyll b (B) and carotenoids (C) contents in leaves of five halophytic plants. Open bars (location 1 far from the factory) and shaded bars (location 2 near to the factory) subjected to dust pollution. Vertical bars represent standard errors (n=3). Values significantly different from non polluted in the same plant type according to F-test ( $p < 0.05$ ).

Table 2: Chlorophyll a/b, total chlorophyll/ carotenoids, soluble sugars/ total sugar and soluble protein/ total protein ratios in five halophytic plants subjected to dust pollution in two localities. Location 1 (loc.1) control far from the factory, location 2 (loc.2) stressed near the factory.. Values are the mean of five readings (n=5).

Plant species	Chl a/b		Total Chl./ Carotenoids		Soluble sugar/ Total sugars		Soluble protein/ Total protein	
	Loc. 1	Loc. 2	Loc. 1	Loc. 2	Loc. 1	Loc. 2	Loc. 1	Loc. 2
<i>Zygophilum coccineum</i>	3.1 (100%)	2.5 (84%)	3.74 $\pm$ 0.18 (100%)	2.3 $\pm$ 0.15 (62%)	0.49 $\pm$ 0.08 (100%)	0.70 $\pm$ 0.11 (143%)	0.44 $\pm$ 0.10 (100%)	0.48 $\pm$ 0.09 (109%)
<i>Salsola tertandra</i>	2.5 (100%)	2.86 (114%)	6.00 $\pm$ 0.32 (100%)	5.31 $\pm$ 0.29 (88.5%)	0.14 $\pm$ 0.008 (100%)	0.18 $\pm$ 0.06 (128%)	0.26 $\pm$ 0.09 (100%)	0.31 $\pm$ 0.009 (119%)
<i>Cyperus conglomeratus</i>	1.8 (100%)	3.4 (189%)	4.3 $\pm$ 0.20 (100%)	4.98 $\pm$ 0.20 (100%)	0.09 $\pm$ 0.005 (100%)	0.12 $\pm$ 0.05 (133%)	0.12 $\pm$ 0.06 (100)	0.16 $\pm$ 0.07 (133%)
<i>Limonium axillare</i>	2.5 (100%)	2.15 (86%)	4.70 $\pm$ 0.17 (100%)	4.40 $\pm$ 0.20 (107%)	0.10 $\pm$ 0.006 (100%)	0.31 $\pm$ 0.08 (310%)	0.12 $\pm$ 0.004 (100%)	0.35 $\pm$ 0.09 (291%)
<i>Suaeda vermiculata</i>	2.7 (100%)	2.10 (79%)	5.01 $\pm$ 0.19 (100%)	4.6 $\pm$ 0.27 (111%)	0.20 $\pm$ 0.06 (100%)	0.32 $\pm$ 0.009 (160%)	0.13 $\pm$ 0.007 (100%)	0.26 $\pm$ 0.01 (200%)

Total carbohydrates content were markedly decreased with increasing dust pollution near the factory for the leaves of *Salsola tetandra* and *Suaede vermiculata* to less than 16 and 29%, respectively compared with non polluted plants, whereas, increased significantly near to the factory and parallel to dust accumulation in *Zygophilum coccineum*, *Cyperus conglomerotus* and *Limonium axillare*. Considerable increases in the soluble sugars that were detected in all plants except *Suaede vermiculata*. Similarly, total protein increment was recorded, especially the soluble protein content in all polluted plants near to the factory except *Salsola tetandra*. Soluble/total protein and soluble/ total carbohydrates ratios were increased in all polluted plants compared with the non polluted plants (Table2). The comparison between the accumulation levels of Fe, Mn, Mg and N in the leaves of polluted plants and control conditions are shown in (Fig.4). The accumulated elements in leaves of plants differed from one species to another in response to cement dust pollution. All the elements (Fe, Mn, Mg and N) in the leaves of *Z. coccineum* and *S. tetandra* were significantly lower in the polluted plants than in the control plants, whereas, their contents in the leaves of *C. conglomerotus* and *S. vermiculata* were significantly higher. In leaves of *L. axillare*, the level of element's accumulation (Mg and N) was significantly lower in polluted plant than in control plant but significantly it was higher for Mn and Fe (Fig.4).

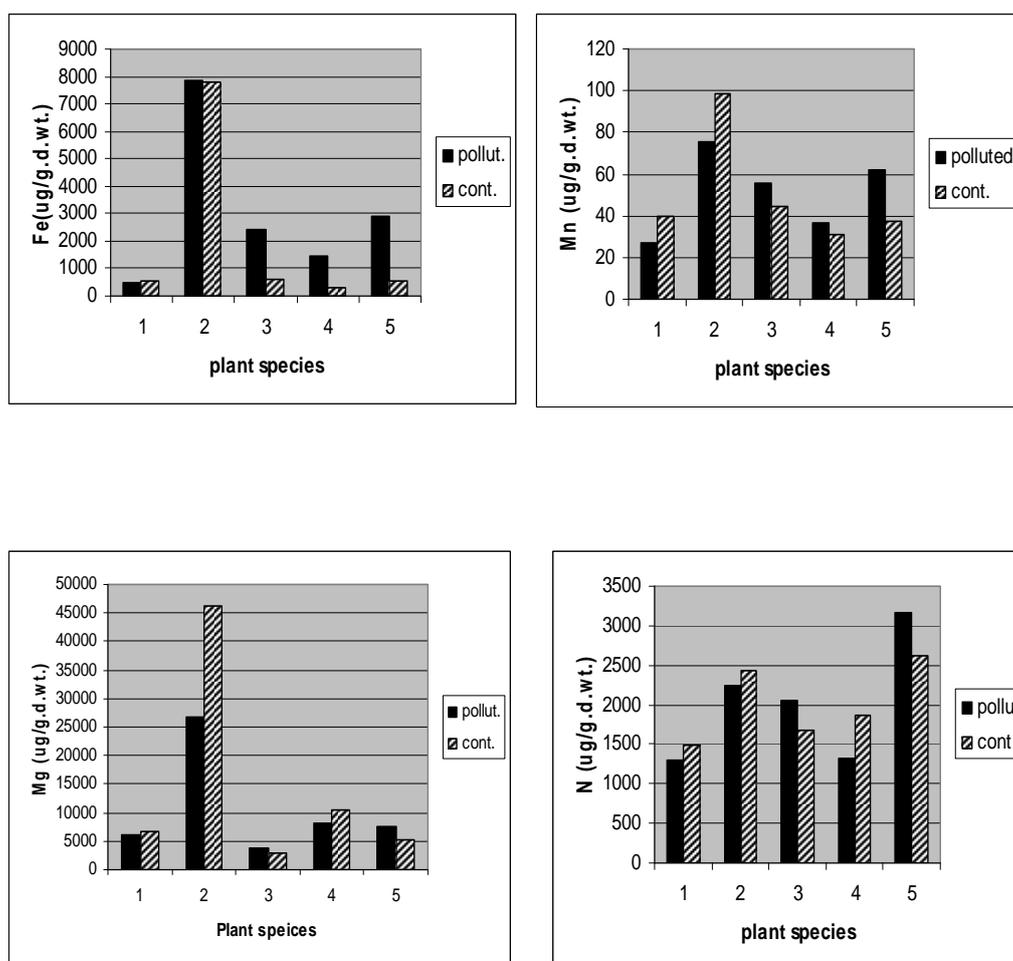


Fig.4 : Element content Fe, Mn, Mg and N in leaves of five halophytic plants 1, *Zygophyllum coccineum*; 2, *Salsola tertandra*; 3, *Cyperus conglomerotus*; 4, *Limonium axillare* and 5, *Suaede vermiculata* in control plants away from the cement dust and polluted plants near to the cement factory. Values significantly different from non polluted in the same plant type according to F-test ( $p < 0.05$ ).

The efflux of electrolytes and pH in all species extracts increased in response to dust pollution and significantly increased in parallel to the dust accumulation in comparison to the control (Fig.5). The present results showed that the two locations (polluted and control) received different amounts of cement dust as indicated by the measurements of pH for the leaves' washing solution. The pH ranged in polluted plant from 8.0 - 8.6 and in control site from 6.7 - 7.2), whereas, the accumulation of the cement dust increased the alkalinity of the cell sap for the plant leaves near the factory (nearly 9-9.1). The polluted soil had a significantly higher pH and electrolyte conductivity (E.C.; 9.2, 2.7) than in the control soil (7.87, 1.6), respectively (Fig. 5).

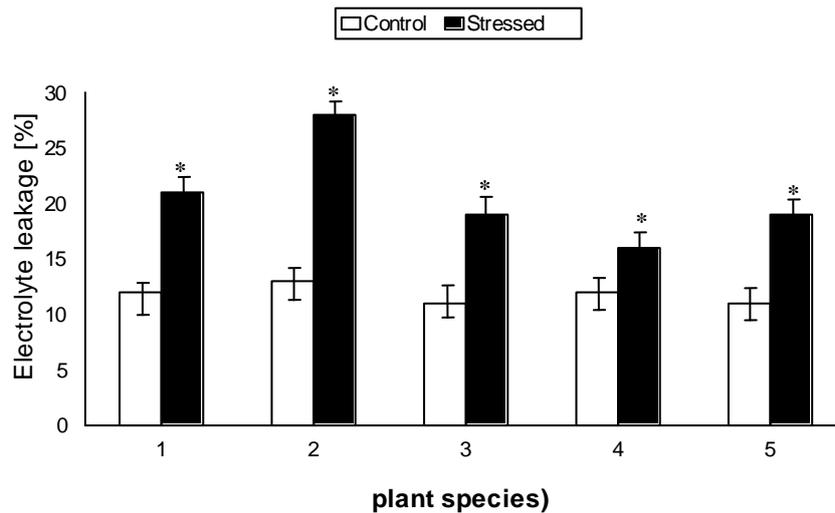


Fig. 5: Electrolyte leakage in leaves of five halophytic plants 1, *Zygophilum coccineum*; 2, *Salsola tetandra*,; 3, *Cyperus conglomerotus*; 4, *Limonium axillare* and 5, *Suaeda vermiculata* in control plants far from the cement dust and polluted plants near to the cement factory. Values significantly different from non polluted in the same plant type according to F-test ( $p < 0.05$ ).

## DISCUSSION

Cement dust had a significant effect on the growth of some plant species compared with non cement dusted plants. Toxic compounds such as fluoride, magnesium, lead, zinc, copper, beryllium, sulfuric acid and hydrochloric acid were found to be produced by cement manufacturing plants (Andrzej, 1987). The results obtained are in close conformity with those reported by Stratmann VH, Van Haut (1969) in which dusted plants with dust ranging from 1 to 48 g/m<sup>2</sup> day<sup>-1</sup> and concluded that dust falling on the soil caused a shift in pH to the alkaline side, which was unfavorable to oats but favorable to pasture grass. Our results are also confirmed by the findings of Prasad and Inamdar (1990) in studying of *Vigna mungo* (Black gram). A quantitative estimation along with dust analysis confirms the ill effects of the cement dust on the plant growth. The pH of dust sample was alkaline in nature; its pH was 9.5 and could be correlated with the findings of Lerman and Darley (1975). They also found a similar range (9.5-11.5) of cement dust pH. The changes in the accumulation of mineral plant nutrients as a result of cement dust were also determined and showed a consistent result with those obtained by Lal and Ambash (1981). It is also suggested that the complete analysis of the cement dust contains all the toxic pollutants should be carried out in detail. The heavy metals present in the cement dust may play an important role in disturbing the various metabolic processes. As it would be possible to recommend these plants in the current study for use as screen or green belts in the industrial localities and adverse urban localities in order to

mitigate dust and improve air quality (Yunas *et al.*, 1985). Dust analysis indicated the presence of heavy metals which may disturb the physiological and some other biochemical reactions. Our results are consistent with the results of George and Ilias (2007) who reported that the traces of toxic metals such as chromium and copper are common in some varieties of Portland cement and are harmful to human being and other living systems. Our data indicates that the exposure of plants to dust altered several physiological and biochemical parameters that were triggered. The most apparent effect of stress induced by dust, described in numerous species, is leaf damage (Naidoo and Chirkoot, 2004). Heath and Castillo, 1988 reported that leaf injury is due to diverse alterations at the sub cellular level .Various studies have shown that the main detrimental effect of dust at the subcellular level is photosystem damage (Nanos and Ilias, 2007; Santosh and Tripathi, 2008). Moreover, the presented results clearly showed that dust altered several biochemical aspects, such as pigment and carbohydrate amount in leaves. A significant percentage increase in chl *a/b* ratio was observed only in *Cyperus conglomeratus* which was mainly caused by an increase in chlorophyll *a* content associated with decrease in chlorophyll *b* content. On the other hand, a marked percentage decrease in chl *a/b* ratio was observed in the other four studied plants. The changes in chlorophyll *a* and *b* are possibly due to shading and/or photosystem damage due to dust accumulation between the peltates or other effects on stomata. Dust from a cement factory seems to cause substantial changes to leaf physiology, possibly leading to reduced plant productivity. Our results are consistent with Nanos and Ilias, (2007) who reported that cement dust decreased the leaf total chlorophyll content and chlorophyll *a*/chlorophyll *b* ratio. As a result, photosynthetic rate and quantum yield decreased. The decrease in the chlorophyll/carotenoid ratio for stressed plants suggested that these relationships could be used as an indicator of tolerance and physiological status of the plants under these stress conditions. According to the results chl *a/b* ratio were significantly increased in *Cyperus conglomeratus* and correlated positively with dust accumulation near to the factory, indicating the presence of protection mechanism of the chloroplast towards the dust pollution. (Lichtenthaler, 2000) stated that the increase in the ratio Chl *a/b* is always associated with a change in pigment composition of the photosynthesis apparatus towards a more sun-type like chloroplast which possesses less light harvesting chlorophyll proteins (LHCPs). According to our results, chloroplasts, in most studied polluted plant, are known to also compose a higher carotenoid content on a chlorophyll basis than non polluted chloroplasts, which is indicated by the lower values for chlorophylls (*a+b*) to carotenoids ratios. In addition, carotenoids, increased in the dust-stressed plants, not play a role as accessory light-harvesting pigments only but they also protect photosynthetic systems against reactive oxygen species (Young, 1991; Asada *et al.*, 1998; Loggini *et al.*, 1999). Our results are consistent with results of (Lichtenthaler *et al.*, 1981, 1982; Lichtenthaler HK, Burkart, 1999) who studied the response of many plants toward various stresses. Moreover, Young, 1991 reported that this response was induced, not only by changes in the irradiation under which the plants are grown, but also by diverse chemicals or stressors that are often associated with long term stress. Carbohydrates content was increased in the leaves of dust polluted plants. The significant increase in the carbohydrates content seems to be involved in osmotic adjustment (Wardlaw IF, Willenbrink, 1994; Tabaeizadeh, 1998). The observed increase in the concentration of soluble sugars may be attributed to growth inhibition by dust stress than photosynthesis, increasing partitioning of fixed carbon to sucrose, (Chaves, 1991; Quick *et al.*, 1992), osmoregulation and tolerance contributing plant survival (Hare *et*

*al.*, 1998). The large alterations observed in plant sugar metabolism preceded the drastic decrease of soluble leaf protein. These proteins are typically related to stress responses, such as freezing, osmotic and salt stress and pathogen attack (Chen *et al.*, 1994; Yun *et al.*, 1996; , Moons *et al.*, 1997; Riccardi *et al.*, 1998; Trudel *et al.*, 1998). Thus the response of studied plants seems to have characteristics in common to dust pollution and in agreement with suggestions made for other species (Shinozaki and Shinozaki, 1996; Tabaeizadeh, 1998). In conclusion, the comparison of five species of different dust susceptibility permitted determining that the tolerance or sensitivity to dust pollution was clearly manifested throughout the photosynthetic activity. The exposure to dust pollution stress provoked important reductions in photosynthesis in most studied plants except *Cyperus conglomeratus*. This study indicates that exposure to particulate deposition may alter plant growth without physical damage to the plant. Moreover, accumulation of dust particulates on studied halophytic leaves could be a major problem in their production. We propose that the pigments content of the light harvesting complex is an important aspect related to the tolerance of plants to dust pollution.

#### ACKNOWLEDGEMENT

The authors extend their appreciation to the Research Center of Teachers College, King Saud University for funding this work through the project.

#### REFERENCES

- Allen, S., H. Grimshay, J. Parkinson and C. I. Quarmby. 1974. *Chemical Analysis of Ecological Materials*. Blackwell Scientific Publications. Osney. Oxford. London.pp.565.
- Andrzej, J. 1987. Bees and their products as indicators of environment pollution. *Med Water*. 43(6): 352–356.
- Armbrust, D. V. 1986. Effect of Particulates (Dust) on Cotton Growth, Photosynthesis, and Respiration. *Agron. J.* 78:1078-1081.
- Asada, K., T. Endo, J. Mano and C. Miyake. 1998. *Molecular mechanism for relaxation of and protection from light stress* In: K. Saton and N. Murata, Editors, Stress responses of photosynthetic organisms, Elsevier, Amsterdam. pp. 37–52.
- Bader, N. B., A. El-Fiky, A. Mustafa and B. Al-Mur. 2009. Metal pollution records in core sediments of some Red Sea coastal areas, Kingdom of Saudi Arabia. *Environmental Monitoring and Assessment*. 155: 1-4.
- Berry, F. and M. B. Bjorkman. 1980. Photosynthetic adaptation to temperature. General features of photosynthetic carbon assimilation versus temperature. *Plant Eco-physiology*, lecture topics 6  
<http://ib.berkeley.edu/courses/ib151/IB151Lecture6.pdf>
- Chaves, M. M. 1991. Effects of water deficits on carbon assimilation. *J. Exp. Botany*. 42: 1–16.
- Chen, R. D., L. X. Yu, A. F. Greer, H. Cheriti and Z. Tabaeizadeh. 1994. Isolation of an osmotic stress- and abscisic acid-induced gene encoding an acidic endochitinase from *Lycopersicon chilensis*. *Mol. Genes and Genetics*. 245: 195-202.
- Dicagno, R., L. Guidi, A. Stefani and F. Soldatini. 1999. Effects of cadmium on growth of *Helianthus annuus* seedlings. *New Phytol.* 144: 65-71.

- George, D. N. and F. I. Ilias. 2007. Effects of inert dust on olive (*Olea europaea* L.) leaf physiological parameters. *Environmental Science and Pollution Research*. 14(3): 212-214.
- Green, N. 2004. *Planning Application for Concrete Batching Plant*. Cranford Way, Hornsey, Application No. HGY/2004/1265. www.GreenN8.org.
- Hare, P.D., W. A. Cress and J. Staden. 1998. Dissecting the roles of osmolyte accumulation during stress. *Plant, Cell and Envir.* 21: 535–553.
- Hartree, E. F. 1972. A modification of Lowery method that gives a linear photometric response. *Analyt. Biochem.* 48:422.
- Heath, R.L. and F. J. Castillo. 1988. *Membrane disturbances in response to air pollutants*, in: S. Schulte-Hosted, L. Blank, N. Darrall, A.R. Wellburn (Eds.), *Air Pollution and Plant Metabolism*, De Gruyter, Berlin, pp. 55–75. Jackson M L (1962). *Soil chemical analysis*. Constable and Comp. Ltd. England.
- Jacobson, J.S. and A. C. Hill. 1970. *Recognition of Air Pollution Injury to Vegetations: A Pictorial Atlas*. Air Pollution Control Association Pittsburgh. Pennsylvania.
- Kilmer, V.J. and L. T. Alexander. 1949. Methods of makings mechanical analysis of soils. *Soil Sci.* 68: 15-24.
- Lal, B. and R. S. Ambasht. 1981. Impact of cement on the mineral and energy concentration of *Psidium guayava*. *Environmental Pollution (Series A)*. 29: 241–249.
- Lerman, S. L. and E. F. Darley. 1975. *Particulates, In. Responses of Plants to Air Pollution*. pp. 141. New York: Academic Press.
- Lichtenthaler, H.K., F. Babani, G. Langsdorf and C. Buschmann. 2000. Measurement of differences in red chlorophyll fluorescence and photosynthetic activity between sun and shade leaves by fluorescence imaging. *Photosynthetica* 38: (4) 521–529.
- Lichtenthaler, H.K., C. Buschmann, M. Döll, H. J. Fietz, T. Bach, U. Kozel, D. Meier and U. Rahmsdorf. 1981. Photosynthetic activity, chloroplast ultrastructure, and leaf characteristics of high-light and low-light plants and of sun and shade leaves, *Photosynth. Res.* 2 : 115–141.
- Lichthenthaler, H. K., G. Kuhn, U. Prenzel, C. Buschmann and D. Meier. 1982. Adaptation of chloroplast-ultrastructure and of chlorophyll-protein levels to high-light and low-light growth conditions, *Z Naturforsch.* 37: 464–475.
- Lichthenthaler, H.K. and S. Burkart. 1999. Photosynthesis and high light stress, *Bulg. J Plant Physiol.* 25: (3-4) 3–16.
- Loggini, A., E. Scartazza, A. Brugnoli and F. Navari-Izzo. 1999. Antioxidant defense system, pigment composition and photosynthetic efficiency in two wheat cultivars subjected to drought, *Plant Physiol.* 119: 1091–1099.
- Metzner, H., H. J. Rouh and H. Senger. 1965. Untersuchungen zur Synchronisierbarkeit einzelner Pigment mangelmutanten von *Chlorella*. *Planta*. 65: 186-194.
- Miller, R.J., J. E. Bittell and D. E. Koeppe. 1973. The effect of cadmium on electron and energy transfer reactions in corn mitochondria, *Physiologia Plantarum* 28: 166-171.
- Moons, A., E. Prisen, G. Bauw and M. Van Montagu. 1997. Antagonistic effects of abscisic acid and jasmonates on salt stress-inducible transcripts in rice roots. *The Plant Cell*. 9: 2243–2259.
- Murata, T., T. Akazawa and F. Shikiko. 1968. Enzymic mechanism of starch breakdown in germinating rice seeds. *Plant Physiol.* 43:1899-1905.

- Naguib, M. I. 1964. Effect of seven on the carbohydrate and nitrogen metabolism during the germination of cotton seeds. *Ind. J. Exp. Biol.* 2:149.
- Naidoo, G. and D. Chirkoot. 2004. The effects of coal dust on photosynthetic performance of the mangrove, *Avicennia marina* in Richards Bay, South Africa. *Environmental Pollution*. 127 (3), : 359-366
- Nanos, G.D. and I. F. Ilias. 2007. Effects of inert dust on olive (*Olea europaea* L.) leaf physiological parameters. *Environ Sci. Pollut. Res. Int.* 14(3):212-4.
- Prasad, M.S.V. and J. A. Inamdar. 1990. Effect of cement kiln dust pollution on ground nut. *Indian Bot Cont.* 7(4):159–162.
- Quick, W.P., M. M. Chaves, R. Wendler, D. David, M. L. Rodrigues, J. Passarinho, J. S. Pereira, M. D. Adcock, R. C. Leegood and M. Stitt. 1992. The effect of water stress on photosynthetic under field conditions. *Plant, Cell and Environ.* 15:25–35.
- Riccardi, F., P. Gazeau, D. de Vienne and M. Zivy. 1998. Protein changes in response to progressive water deficit in maize. *Plant Physiol.* 117: 1253–1263.
- Richards, L. A. 1954. Diagnosis and improvement of saline and alkali soils. USDA Handbook No. 60.
- Santosh, K.P. and B. D. Tripathi. 2008. Seasonal Variation of Leaf Dust Accumulation and Pigment Content in Plant Species Exposed to Urban Particulates Pollution. *J Environ Qual.* 37:865-870.
- Schutzki, R.E. and B. Cregg. 2007. Abiotic Plant Disorders Symptoms, Signs and Solutions. A Diagnostic Guide to Problem Solving. *Extension Bulletin.* E-2996.
- Shinozaki, K. and K. Y. Shinozaki. 1996. Molecular responses to drought and cold stress. *Current Opinion in Biotech.* 7:161–167.
- Stratmann, V.H. and H. Van Haut. 1966. In: Darley EF. Studies on the effect of cement kiln dust on vegetation. *Air Pollution Control Association Journal.* 165(3): 33–39.
- Tabaeizadeh, Z. 1998. Drought-induced responses in plant cells. *Inter. Rev. of Cytology.* 182: 193–247.
- Taylor, G. and W. J. Davies. 1990. Root growth of *Fagus sylvatica*: Impact of air quality and drought at a site in southern Britain. *New Phytologist.* 166: 457-464.
- Thomas, P. E. 1969. *The analysis of cement.*  
<http://big5.varianinc.com.cn/products/spectr/aa/articles/aa-01.pdf>
- Trudel, J., J. Grenier, C Potvin and A. Asselin. 1998. Several thaumatin-like proteins bind to  $\beta$ -1,3-glucans. *Plant Physiol.* 118: 1431–1438.
- Wardlaw, I.F. and J. Willenbrink. 1994. Carbohydrate storage and mobilization by the culm of wheat between heading and grain maturity: the relation to sucrose synthase and sucrose-phosphate synthase. *Aust. J. Plant Physiol.* 21: 255–271.
- Young, A. 1991. The photoprotective role of carotenoids in higher plants, *Physiol Plant.* 83: 702–708.
- Yun, D.J., M.P. D'Urzo, L. Abad, S. Takeda, R. Salzman, Z. Chen, H. Lee, P.M. Hasegawa and R. A. Bressan . 1996. Novel osmotically induced antifungal chitinases and bacterial expression of an active recombinant isoform. *Plant Physiol.* 111: 1219–1225.
- Yunas, M., A.K. Dwivedi, K .Kulshreshtha and K.J. Ahmed. 1985. Dust loading on some common plants near Lucknow City. *Environmental Pollution (Series B).* 9: 7–80.